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Two new phenylpropanoids and one propanoate from *Morinda citrifolia*

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Two new phenylpropanoids, methyl 3-(2,4-dihydroxy-5-methoxyphenyl)propionate (**1**) and butyl 3-(2,4-dihydroxy-5-methoxyphenyl)propionate (**2**), and one unusual propanoate, 5-hydroxyhexyl 2-hydroxypropanoate (**3**), were isolated from the fruits of *Morinda citrifolia*. Their structures were established using MS and NMR methods.

Keywords: *Morinda citrifolia*; Rubiaceae; phenylpropanoids; propanoate

1. Introduction

Morinda citrifolia L. (Rubiaceae), known as ‘noni’, is a small tree that grows widely across Polynesia [1]. There is a long history of the use of *M. citrifolia* as an important medicinal plant for the treatment of asthma, bone fractures, cancer, cholecystitis, dysentery, lumbago, menstrual cramps, urinary difficulties, and many other ailments [2]. Recent phytochemical studies on this plant have reported anthraquinones and anthraquinone glycosides, fatty acid glycosides, iridoids and iridoid glycosides, lignans, and triterpenoids. Of them, the novel iridoids including citrifolinin A [3], citrifolinoside [4], yopaaoside A [5], yopaaoside B [5], and morindacin [6] were isolated. They showed activities in a wide range of biological assays such as the inhibition of angiogenesis, 5-lipoxygenase and 15-lipoxygenase, cyclooxygenase-1, phorbol ester-induced inflammation and

tyrosine kinase [7–11]. Here, we report the isolation of three new compounds, methyl 3-(2,4-dihydroxy-5-methoxyphenyl)propionate (**1**), butyl 3-(2,4-dihydroxy-5-methoxyphenyl)propionate (**2**), and 5-hydroxyhexyl 2-hydroxypropanoate (**3**) (Figure 1), from the EtOAc and *n*-BuOH extract of the fruits of *M. citrifolia*.

2. Results and discussion

Compound **1** was isolated as a yellow gum. The molecular formula C₁₁H₁₄O₅ was established by the positive HR-ESI-MS at *m/z* 249.0745 [M + Na]⁺. In the ¹H NMR spectrum, two singlets at δ_H 3.81 and 3.68 (each 3H) were assigned to two methoxyl groups, two singlets at δ_H 6.55 and 6.52 (each 1H) were assigned to two aromatic protons, and four protons at δ_H 2.82 and 2.68 (each 2H, t, *J* = 6.4 Hz) were ascribed to protons of two conjoint methylenes. The ¹³C NMR spectrum displayed 11 carbon resonances ascribable to 2 methoxyl groups

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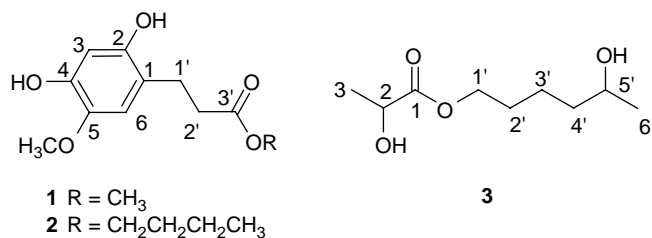


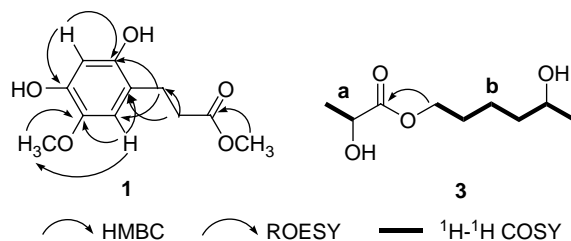
Figure 1. Chemical structures of compounds 1–3.

(δ_C 56.5, 52.3), 2 methylenes (δ_C 23.5, 29.4), 2 methines (δ_C 104.0, 109.6), and 5 quaternary carbons (δ_C 113.0, 138.6, 144.5, 147.4, 169.3). These NMR spectral data suggested that compound **1** was an α,β -saturated phenylpropanoid. One of the methoxyl groups was placed at C-5 according to the HMBC correlation between 5-OCH₃ and C-5, and the ROESY correlation of H-6 with 5-OCH₃ (Figure 2). In addition, the HMBC correlation of the methoxyl at δ_H 3.68 (3H, -OCH₃) with C-3' at δ_C 169.3 suggested a methyl ester group (Figure 2). Detailed analysis of HMBC and ROESY spectra (Figure 2) established the structure of compound **1** as methyl 3-(2,4-dihydroxy-5-methoxyphenyl)propionate.

Compound **2** was isolated as a yellow gum. The HR-ESI-MS established the molecular formula C₁₄H₂₀O₅ (m/z 291.1201 [M + Na]⁺, calcd 291.1208). All the NMR spectral data suggested that compound **2** had a similar structure to that of **1** except for a *n*-butyl ester in **2** instead of the methyl ester in **1** as suggested by the HMBC correlations of the oxymethylene at

δ_H 4.08 (2H, t, J = 6.5 Hz) with C-3' at δ_C 175.4 and the MS data. Detailed analysis of 2D NMR (HSQC, HMBC, ¹H-¹H COSY) spectra finally established the structure of compound **2** as butyl 3-(2,4-dihydroxy-5-methoxyphenyl)propionate. However, compound **2** might be an artificial product due to the usage of *n*-BuOH in the extraction.

Compound **3** was obtained as a colorless oil. Its molecular formula C₉H₁₈O₄ was determined by the negative HR-ESI-MS at m/z 189.1127 [M-H]⁻. The IR spectrum revealed the presence of hydroxyl (3389 cm⁻¹) and carbonyl (1736 cm⁻¹) groups. The ¹³C NMR spectrum displayed nine carbon resonances including one carbonyl group (δ_C 175.8), two methines (δ_C 66.7, 67.8), four methylenes (δ_C 22.0, 28.5, 38.6, 65.5), and two methyl (δ_C 20.4, 23.6) carbons. Correspondingly, the ¹H NMR spectrum revealed two methyl groups at δ_H 1.19, (d, J = 6.2 Hz) and 1.41 (d, J = 6.2 Hz), two oxymethines at δ_H 4.27 (m) and 3.79 (m), one oxymethylene at δ_H 4.19 (m), and three methylenes at δ_H 1.44–1.68.

Figure 2. Key 2D NMR correlations of **1** and **3**.

The ^1H - ^1H COSY spectrum revealed two partial fractions (**a** CH_3CH and **b** $\text{CH}_3\text{CHCH}_2\text{CH}_2\text{CH}_2\text{CH}_2$) (Figure 2). In the HMBC spectrum, the key correlation of an oxymethylene at δ_{H} 4.17 (2H, m, H-1') with C-1 at δ_{C} 175.8 suggested the connection of partial fractions **a** and **b** via through an ester group (Figure 2). Thus, the planar structure of **3** was determined as 5-hydroxyhexyl 2-hydroxypropanoate. Unfortunately, the stereo configuration at C-2 and C-5' could not be identified due to the limited quantity available.

3. Experimental

3.1 General experimental procedures

Optical rotations were measured with a Horiba SEPA-300 polarimeter. UV spectra were obtained using a Shimadzu UV-2401A spectrophotometer. A Tenor 27 spectrophotometer was used for scanning IR spectroscopy using KBr pellets. 1D and 2D NMR spectra were run on Bruker DRX-500 and AM-400 spectrometers with TMS as the internal standard. Chemical shifts (δ) were expressed in ppm with reference to the solvent signals. HR-ESI-MS were performed on an API-Qstar-Pulsar-1 spectrometer. Column chromatography was performed on silica gel (200–300 mesh, Qingdao Haiyang Chemical Co. Ltd., Qingdao, China) and RP-18 gel (20–45 μm , Fuji Silysia Chemical Ltd., Aichi, Japan). Fractions were monitored by TLC (GF 254, Qingdao Haiyang Chemical Co. Ltd.), and spots were visualized by heating silica gel plates sprayed with 10% H_2SO_4 in EtOH.

3.2 Plant material

The dried powder of the fruit of *Morinda citrifolia* was obtained from Hsiehs Biotech., Vietnam in March 2008 and identified by Prof. Hua Peng, and a voucher specimen (YP08019) has been deposited in Hsiehs Biotech., Vietnam.

3.3 Extraction and isolation

Dried noni fruits (1100 g) were extracted with 95% EtOH (11) at room temperature for 1 week. The extract was concentrated to dryness under reduced pressure, and the residue was suspended in H_2O (1000 ml) and partitioned successively with EtOAc (3×500 ml) and *n*-BuOH (3×500 ml). The *n*-BuOH-soluble fraction (61 g) was subjected to column chromatography over silica gel (600 g, 140 cm \times 7 cm; CHCl_3 : $\text{Me}_2\text{CO} = 1:0 \rightarrow 1:1$) to give six subfractions 1–6. Subfraction 5 (4 g) was separated by RP-18 (100 g, 60 cm \times 4 cm; $\text{MeOH}:\text{H}_2\text{O} = 1:1$) to afford **1** (1.1 mg) and **2** (2.5 mg). The EtOAc fraction (21 g) was subjected to column chromatography on silica gel column chromatography (400 g, 60 cm \times 4 cm; CHCl_3 : $\text{Me}_2\text{CO} = 1:0 \rightarrow 1:1$) to afford subfractions 6–9. Subfraction 9 (3 g) was separated by silica gel (100 g, 50 cm \times 3 cm; petroleum ether: $\text{Me}_2\text{CO} = 2:1$) and further purified by Sephadex LH-20 (30 g, 150 cm \times 2 cm; CHCl_3 : $\text{MeOH} = 1:1$) to yield **3** (2 mg).

3.3.1 Methyl 3-(2,4-dihydroxy-5-methoxyphenyl)propionate (**1**)

Yellow gum; UV (MeOH) λ_{max} (log ϵ): 204 (3.76), 290 (3.02) nm; IR (KBr) ν_{max} : 3439 (OH), 1737 (C=O) cm^{-1} ; ^1H NMR (CDCl_3 , 20°C, TMS) and ^{13}C NMR (CDCl_3 , 20°C, TMS) spectral data, see Table 1. HR MS ((+)-ESI): m/z 249.0745 [$\text{M} + \text{Na}$] $^+$ (calcd for $\text{C}_{11}\text{H}_{14}\text{O}_5\text{Na}$, 249.0738).

3.3.2 Butyl 3-(2,4-dihydroxy-5-methoxyphenyl)propionate (**2**)

Yellow gum; UV (MeOH) λ_{max} (log ϵ): 204 (3.78), 290 (3.16) nm; IR (KBr) ν_{max} : 3441 (OH), 1737 (C=O) cm^{-1} ; ^1H NMR (CDCl_3 , 20°C, TMS) and ^{13}C NMR (CDCl_3 , 20°C, TMS) spectral data, see Table 1. HR MS ((+)-ESI): m/z 291.1201 (calcd for $\text{C}_{14}\text{H}_{20}\text{O}_5\text{Na}$, 291.1208).

Table 1. ^1H and ^{13}C NMR spectral data of **1** and **2** in CDCl_3 (δ in ppm, J in Hz).

Position	1		2	
	δ_{H}	δ_{C}	δ_{H}	δ_{C}
1		113.0 (s)		118.0 (s)
2		147.4 (s)		148.6 (s)
3	6.52 (1H, s)	104.0 (d)	6.52 (1H, s)	104.5 (d)
4		138.6 (s)		140.0 (s)
5		144.5 (s)		145.0 (s)
6	6.55 (1H, s)	109.6 (d)	6.55 (1H, s)	112.6 (d)
1'	2.82 (2H, t, 6.4)	23.5 (t)	2.81 (2H, t, 6.4)	24.2 (t)
2'	2.68 (2H, t, 6.4)	29.4 (t)	2.67 (2H, t, 6.4)	35.5 (t)
3'		169.3 (s)		175.4 (s)
5-OCH ₃	3.81 (3H, s)	56.5 (q)	3.82 (3H, s)	56.5 (q)
3'-OCH ₃	3.68 (3H, s)	52.3 (q)		
OCH ₂ CH ₂ CH ₂ CH ₃			4.08 (2H, t, 6.5)	65.3 (t)
OCH ₂ CH ₂ CH ₂ CH ₃			1.58 (2H, m)	30.4 (t)
OCH ₂ CH ₂ CH ₂ CH ₃			1.33 (2H, m)	19.0 (t)
OCH ₂ CH ₂ CH ₂ CH ₃			0.90 (3H, t, 7.2)	13.6 (q)

3.3.3 5-Hydroxyhexyl 2-hydroxypropanoate (**3**)

Colorless oil; $[\alpha]_{\text{D}}^{20} - 20.1$ ($c = 0.49$, acetone). IR (KBr) ν_{max} : 3389 (OH), 1736 (C=O), 1459, 1214, 1134 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3 , 20°C, TMS): δ 4.26 (1H, m, H-2), 4.17 (2H, m, H-1'), 3.79 (1H, m, H-5'), 1.68 (2H, m, H-2'), 1.47 (2H, m, H-4'), 1.45 (2H, m, H-3'), 1.40 (3H, d, $J = 6.2$ Hz, H-3), 1.18 (3H, d, $J = 6.2$ Hz, H-6'). ^{13}C NMR (100 MHz, CDCl_3 , 20°C, TMS): δ 20.4 (q, C-3), 22.0 (t, C-3'), 23.6 (q, C-6'), 28.5 (t, C-2'), 38.6 (t, C-4'), 65.5 (t, C-1'), 66.7 (d, C-2), 67.8 (d, C-5'), 175.8 (s, C-1). HR MS ((-)-ESI): m/z 189.1127 $[\text{M}-\text{H}]^-$ (calcd for $\text{C}_9\text{H}_{17}\text{O}_4$, 189.1126).

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